



Effects of Low Tidal Volume Ventilation in a Murine Model of Ventilator-induced Diaphragmatic Dysfunction

**Cho, Hwa Jin¹, Hyun Woo Kim², Seong Woo
Kang², Do Hyun Yu², In Seok Jeong³**

¹Department of Pediatrics, Chonnam National University Hospital

²College of Veterinary Medicine, Chonnam National University

³Department of Thoracic and Cardiovascular surgery

Chonnam National University Hospital

Introduction

- ✓ **Mechanical ventilation (MV)** is a life-saving procedure in many critically ill patients.
- ✓ Short period of MV has been associated with ventilator-induced lung injury even in healthy lungs.

Wolthuis EK, et al., ANESTHESIOLOGY 2008

✓ **Controlled MV**

- Maintains the diaphragm **at rest**
- Has been implicated in the development of **diaphragmatic dysfunction**
- Not only in animal models but also in humans.

Capdevila X, et al., Intensive Care Med 2003, Gayan-Ramirez G, et al., Intensive Care Med 2003

Jaber S, et al., Intensive Care Med 2005, Le Bourdelles G, et al., Am J Respir Crit Care Med 1994

Sassoon CS, et al., J Appl Physiol 2002, Yang L, et al., Am J Respir Crit Care Med 2002

Ramirez G: Crit Care 2010, Hussain SN, et al., Am J Respir Crit Care Med 2010

Jaber S, et al., Am J Respir Crit Care Med 2011, Levine S, et al., Am J Respir Crit Care Med 2011

Levine S, et al., N Engl J Med 2008

Ventilator-Induced Diaphragmatic Dysfunction (VIDD)

- Play an important role in the **difficulties in weaning critically ill patients from the ventilator.**

Schultz MJ. ANESTHESIOLOGY 2010

- **Multiple mechanisms** appear to be involved in VIDD
 - increase of reactive oxygen species
 - mitochondrial dysfunction
 - inhibition of the insulin-like growth factor pathway
 - activation of different proteolytic systems, such as the calpains and caspase 3
 - Calpains and caspase 3 are ubiquitous nonlysosomal proteases, and their expression may be up-regulated early in the time course of VIDD.

Goll DE, et al. Physiol Rev 2003

Powers SK, et al. J Appl Physiol 2007

- ✓ All of these mechanisms culminate in
 - muscle atrophy
 - injury
 - loss of diaphragmatic force-generating capacity

Jaber S, et al. Am J Respir Crit Care Med 2011

Jaber S, et al. Crit Care 2011

Petrof BJ, et al. Curr Opin Crit Care 2010

Tobin MJ, et al. Ann Intern Med 2010

VIDD according to Tidal Volume

- ✓ **Low tidal volume ventilation (LTVV)**
- Important concept of **lung protective strategy** from ventilator associated lung injury
- However, there was **no evidence** that LTVV has protective effect **on diaphragm**, especially on ventilator induced diaphragmatic dysfunction.

Study Objectives

- ✓ **To evaluate the effects of LTVV** in murine VIDDD model by exploring both histologic and the main protease pathway after MV

MATERIALS AND METHODS

Materials and methods

Healthy male C57/BL6 mice (10-12 weeks, 25-30g)

- 1) Higher tidal volume MV for 6 h (**HTV group**, n=6)
 - TV: 10 μ l/mg BW in HTV
- 2) Lower tidal volume MV for 6 h (**LTV group**, n=6)
 - TV: 6 μ l/mg BW in LTV
- 3) Controls (Control group, n=6).

- ✓ Anesthesia:
 - IP inj of pentobarbital sodium (50mg/kg body weight)
- ✓ Tracheostomy:
 - 22-gauge angiocatheter was inserted
- ✓ Hemodynamic stability
 - 0.05ml RL solution IP every hour
- ✓ Other general care:
 - bladder expression
 - ocular lubrication
 - passive limb movement



Experimental protocol for MV

✓ Small animal ventilator

: Flexivent®, SCIREQ Inc, Canada

✓ Ventilator setting:

CMV mode

FiO₂: 0.21

RR: 150 rpm

PEEP: 3-4 cm H₂O

Tidal volume

– **10ul/mg BW in HTV**

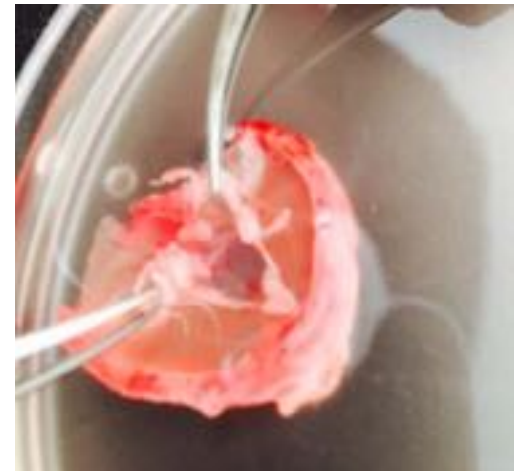
– **6ul/mg BW in LTV**

1. Lung Function Measurement

- In 3 groups
- **Forced oscillation technique in FlexiVent system**
 - Airway resistance [R_n]
 - Tissue damping (resistance)[G]
 - Tissue elasticity [H]

2. Blood collection & Tissue Collection (Diaphragm)

- By thoracotomy and laparotomy



3. Measurement of Diaphragm contractile Properties

✓ The force-frequency relationship

:Enhanced contractility at higher rates of stimulation

- at *10, 20, 30, 50, 60, 80, 100, 120 Hz*
- for *600 ms*
- with 1 min between the stimulation trains.

✓ The fatigability assessment

- measured by loss of force to repeated stimuli
- *over 10 min*
- *with 30 Hz*
- *300 ms* duration

4. Measurement of inflammation and atrophy

- ✓ Stained by **Hematoxylin and Eosin**
- ✓ The complete area was evaluated by a blinded investigator for **invasion of neutrophils and lymphocyte**

5. Measurement of the sarcolemmal injury

- ✓ Exposed to a **fluorescent tracer dye**
 - **Procion orange**; Sigma, St. Louis, MO

6. Biochemical Evaluation

- ✓ Total levels of **calpains 1 and 2**
- ✓ **Immunoblot analysis** was performed.
 - **Calpain 1** : ab 49652 1:1,000 dilution in 3% BSA/TBST, 80KDa; Abcam, Cambridge, MA
 - **Calpain 2** : ab39165 1:1000 dilution with TBST containing 5% skim-milk, 76 KDa; Abcam, Cambridge, MA

Statistical Analysis

- Data are presented as mean \pm standard deviation.
- Kruskal-Wallis test
 - additional analysis between 2 groups was performed using Mann-Whitney test.
- Repeated measure of ANOVA.
 - Differences in muscle contractility force-frequency response and muscle contractility for fatigability between groups
- Significance was established at $P < 0.05$.
- Statistical analysis was performed using SPSS 21.0.

RESULTS

Animal characteristics

Table 1. Animal characteristics of all 3 groups and mean peak inspiratory pressure (PIP), positive end expiratory pressure (PEEP), and tidal volume (TV) in ventilated groups (HTV and LTV).

	HTV (n=6)	LTV (n=6)	Control (n=6)	P value
Body weight (g)	22.0 ± 0.9	21.8 ± 1.1	23.6 ± 2.1	0.273
Ventilator		3.2		
PIP	11.0 ± 0.9	8.9 ± 0.6	-	0.006
PEEP	2.5 ± 1.1	2.9 ± 0.03	-	0.873
TV	0.2 ± 0.007	0.12 ± 0.003	-	0.005

Data are represented as mean ± SE.

HTV = high tidal volume; LTV = low tidal volume; PIP = peak inspiratory pressure; PEEP – positive end expiratory pressure; TV = tidal volume.

Arterial blood gas analyses

	PH	PaO ₂ (mmHg)	PaCO ₂ (mmHg)	HCO ₃ ⁻ (mM)
HTV (n=6)	7.4 ± 0.1	65.2 ± 19.7	21.6 ± 4.4	20.0 ± 5.0
LTV (n=6)	7.4 ± 0.1	84.5 ± 11.6	21.0 ± 2.3	22.6 ± 0.9
Control (n=6)	7.4 ± 0.2	92.4 ± 25.3	19.7 ± 5.9	26.1 ± 3.2

Data are represented as mean ± SE.

HTV = high tidal volume; LTV = low tidal volume

No significant differences were observed between groups.

NS

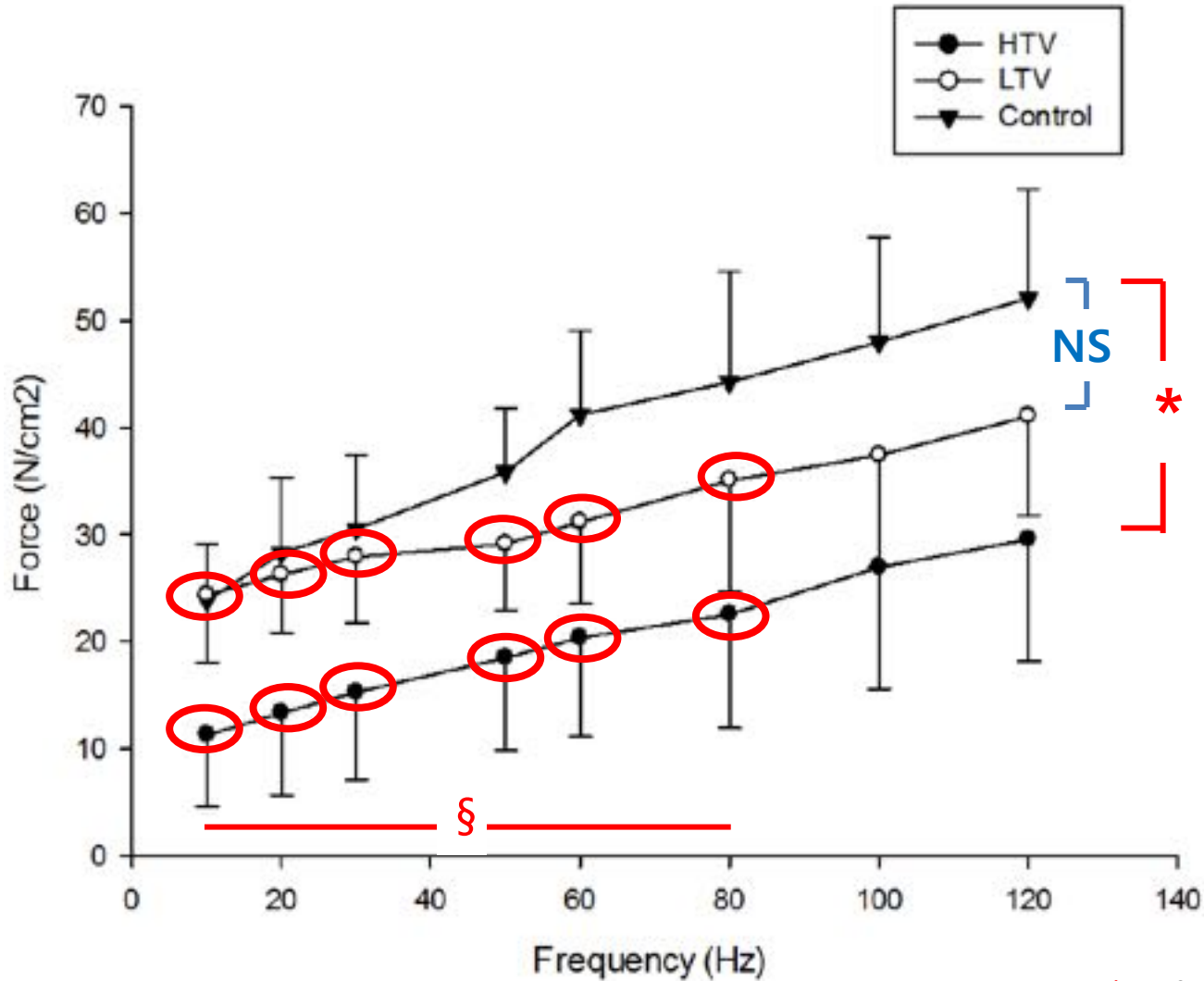
Invasive Lung Function Measurement by Forced oscillation technique (FlexiVent)

	Airway resistance (RN), cmH ₂ O.s/ml	Tissue Damping (G), cm H ₂ O/ml	Tissue Elasticity (H), cm H ₂ O/ml	P value
HTV (n=6)	0.37±0.09	4.48 ± 1.15	33.1 ± 5.79	0.343
LTV (n=6)	0.38 ± 0.05	5.10± 0.71	39.2 ± 4.95	0.158
Control (n=6)	0.32 ± 0.07	4.25 ± 0.77	31.5 ± 7.16	0.087

NS

Force-Frequency Relationship

:Enhanced contractility at higher rates of stimulation

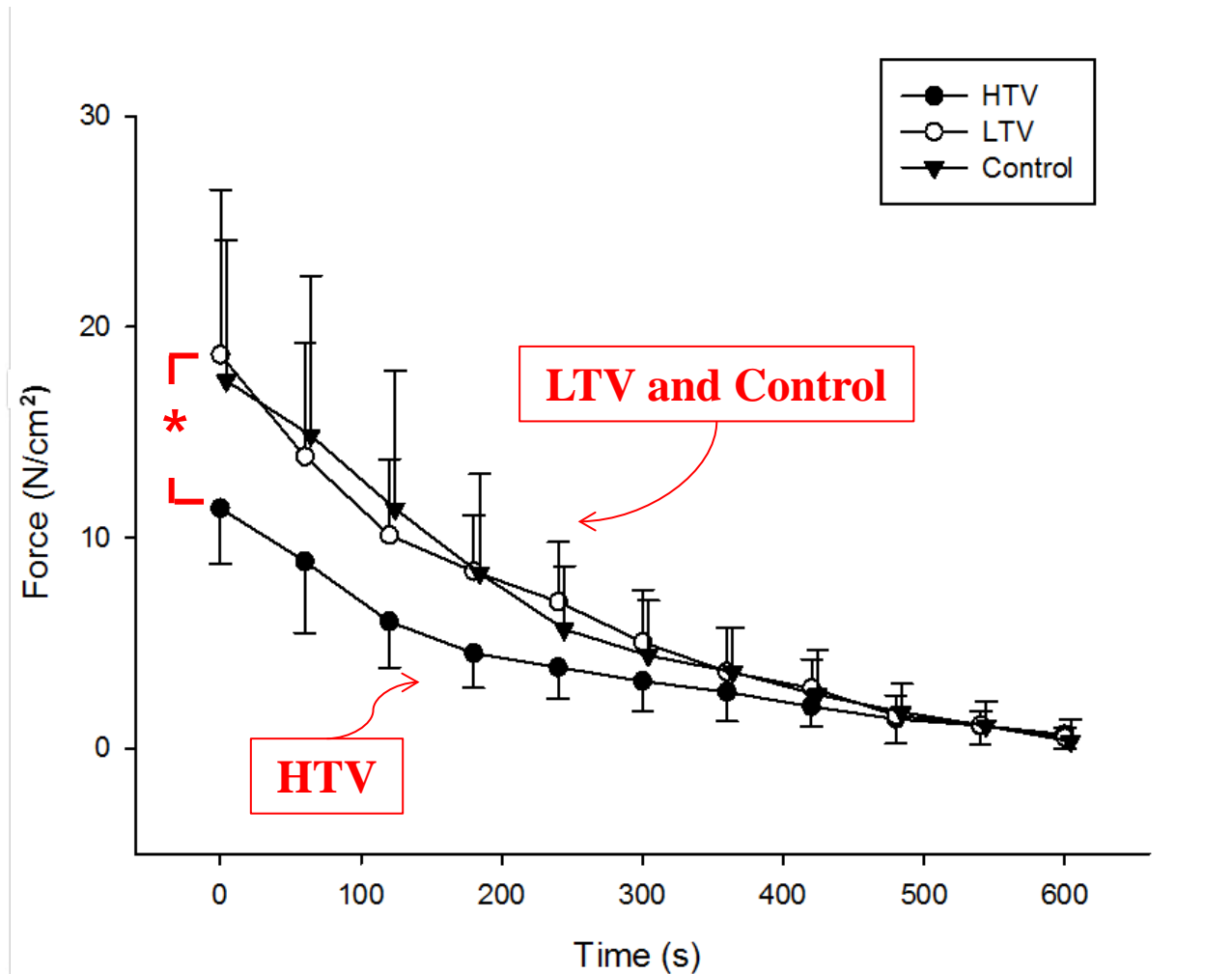


* P<0.05, HTV vs control

§ P<0.05, HTV vs LTV

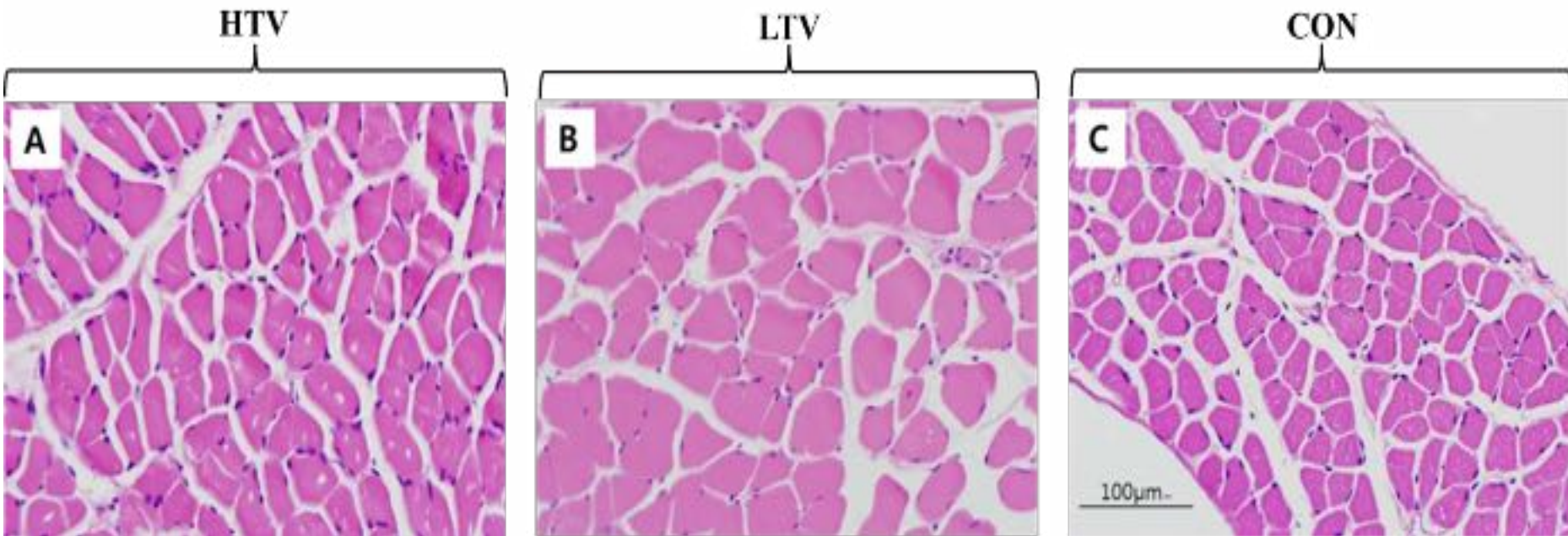
Fatigability Assessment :

Loss of absolute force production of the diaphragm during the fatigue test



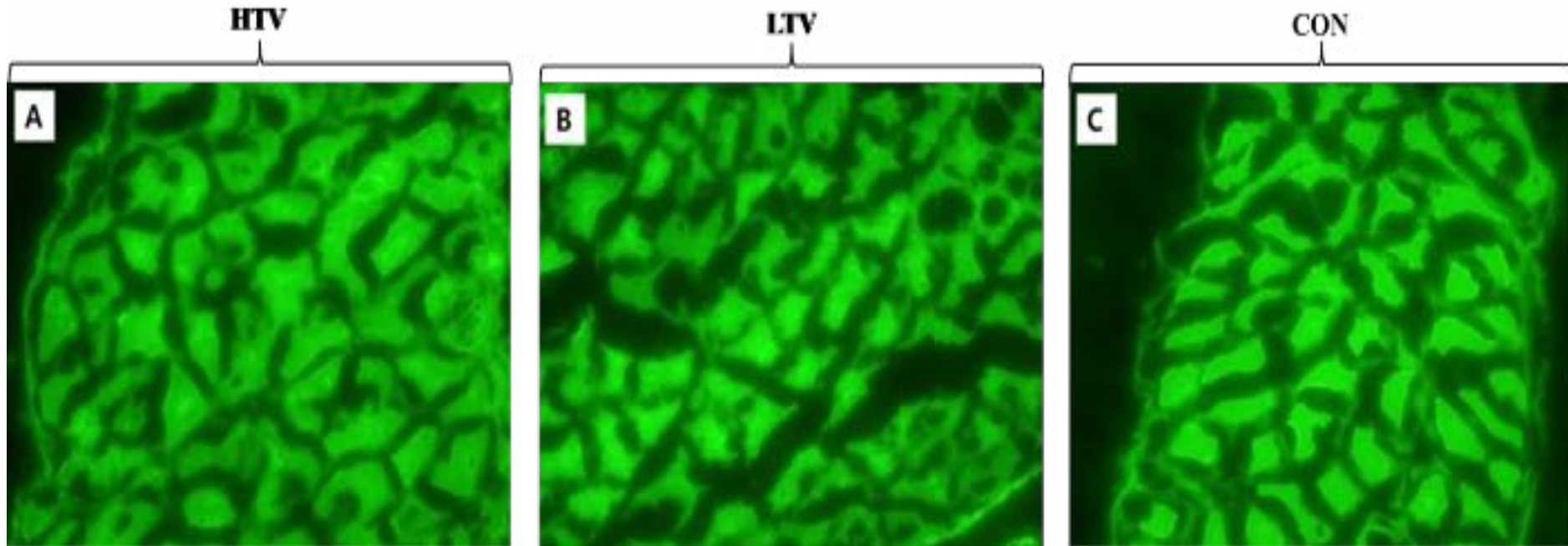
*P<0.05, HTV vs LTV and control groups.

Histologic findings for inflammation in diaphragm



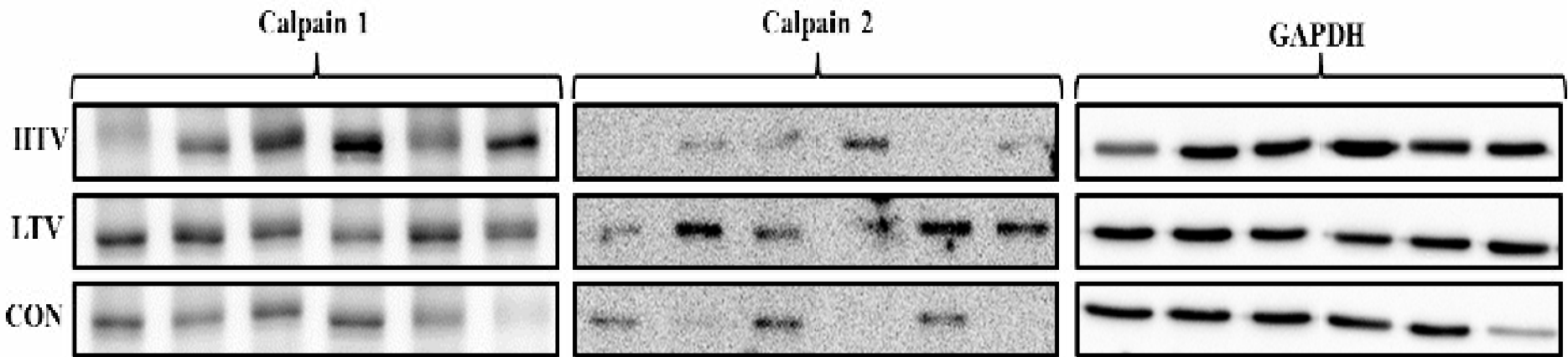
No differences between groups

Quantification of the sarcolemmal injury in diaphragm



No differences between groups

Expression of calpain isoforms in diaphragm



No differences between groups

Conclusion

- ✓ **Low tidal volume ventilation**
 - ✓ **partially attenuates the development of VIDD** in murine model
- ✓ **Reducing ventilatory support to only that level necessary for respiratory system recovery while avoiding harm is just as true for VIDD as it is for VILI.**